

IMMOBILIZATION OF UNMODIFIED BIOPOLYMERS TO ACYL FLUORIDE ACTIVATED SUBSTRATES

Background of the Invention

Area of the Art

The invention relates generally to solid supports with immobilized biopolymers and specifically to solid supports with immobilized unmodified biopolymers and methods of immobilizing unmodified biopolymers to solid supports.

Description of the Prior Art

Biopolymer synthesis and biopolymer analysis often require the attachment of biopolymers to solid supports. For example, organic and inorganic materials have been utilized for the solid phase synthesis of peptides, oligonucleotides and small organic molecules. The synthesis involves the stepwise addition of activated monomers such as amino acid derivatives or nucleotide derivatives to a growing oligomeric chain attached at one end to a solid support. At the completion of the synthesis, the newly synthesized biopolymers may be cleaved from the solid support and subsequently utilized in biochemical research or diagnostic applications or, alternatively, be utilized without cleaving the biopolymers from the solid support.

For biopolymer analysis, biopolymers may be attached to a solid support in several ways. In blotting techniques, native biopolymers are first captured onto a membrane and subsequently immobilized on the membrane by heat, radiation or chemical techniques. The immobilized biopolymers are then available for subsequent analyses, such as those associated with southern blotting applications and reverse hybridization analytical techniques.

Additionally, presynthesized or natural oligonucleotides have been immobilized by covalently attaching activated oligonucleotides to the solid support. Current methodology for the covalent attachment of nucleic acids to solid supports (substrates) involves modification of the DNA (or RNA). For example, oligonucleotides are usually derivatized to a 5'-amino terminus, making the DNA more reactive for covalent attachment to an activated surface. Other methods of attachment have employed reactions with terminal phosphate groups or sulfhydryl groups with surface carbodiimide or other activation chemistries (see Lund et al, *Nucleic Acid Res.* 16: 10861-80, 1988; Bischoff et al, *Analyt. Biochem.* 164: 336-344, 1987).

It is generally understood that reactive groups present within native polynucleotides are weak and therefore make for inefficient attachment. In addition, when native

polynucleotides are exposed to highly reactive surface groups, excessive crosslinking may occur. This crosslink may prevent the attached nucleic acid from fully participating in hybridization. These conditions are most noticeable for short fragments of double-stranded DNA or oligonucleotides. Thus, oligonucleotides often have to be modified, for example, derivatized to a 5'-amino terminus, for an effective attachment. The 5' amino-linker allows selective binding of the amino-containing DNA to silylated slides through a Schiff's base reaction with aldehyde groups on the chip surface. The selectivity of amino-modified versus natural, unmodified DNA is about 10:1 for cDNAs and about 10,100:1 for single-stranded 15-mers. DNA molecules of intermediate lengths exhibit intermediate discrimination ratios. In addition, the 5' end attachment of the DNA to the chip via the amino group permits steric accessibility of the bound molecules during the hybridization reaction. Therefore, post-modification has been perceived as obligatory for attachment of, e.g., oligonucleotide probes for creation of arrays. Such post-modification processes require additional time-consuming steps at substantial costs.

Therefore, it is desirable to develop a more effective method for attaching biopolymers, particularly unmodified biopolymers to a solid support. It is particularly desirable to develop a method to directly attach unmodified biopolymers, such as polynucleotides, to a solid support.

Summary of the Invention

The present invention is based on the discovery that both short and long fragments of single-stranded or double-stranded DNA may be efficiently attached to acyl fluoride activated supports directly without spacer arms and without modifying the polynucleotide. It is also based on the discovery that other biopolymers, such as protein A, antibodies, streptavidin, etc., may also be attached to a solid support without modifications to the biopolymers.

Accordingly, one aspect of the present invention provides a method of attaching unmodified biopolymers to a solid support. The method comprises the steps of:

- (a) providing unmodified biopolymers;
- (b) providing a solid support having at least one surface comprising pendant acyl fluoride functionalities; and
- (c) contacting the unmodified biopolymers with the solid support under a condition sufficient for allowing the attachment of the biopolymers to the solid support.

According to embodiments of the present invention, the biopolymers may be nucleic acids, polypeptides, proteins, carbohydrates, lipids and analogues thereof. In one embodiment of the present invention, the biopolymer is a polynucleotide, and the polynucleotide may be synthesized oligonucleotide, amplified DNA, cDNA, single stranded DNA, double stranded DNA, PNA, RNA or mRNA. Also according to embodiments of the present invention, the solid support may be polymeric materials, glasses, ceramics, natural fibers, silicones, metals and composites thereof.

Another aspect of the present invention provides a method of analyzing a biopolymer target in a sample. The method comprises the steps of:

- (a) providing a solid support fabricated of a material having pendent acyl fluoride groups on at least one surface;
- (b) providing an agent that can form a complex with the biopolymer target, wherein the agent comprises a second biopolymer;
- (c) contacting the solid support with either the agent or the biopolymer target under a condition that allows the attachment of either the unmodified agent or the unmodified biopolymer to the solid support, wherein the agent and the biopolymer target are unmodified;
- (d) contacting the solid support attached with the unmodified agent with the biopolymer target, or contacting the solid support with the attached, unmodified biopolymer target with the agent under a condition that allows the formation of a complex comprising the agent and the biopolymer target;
- (e) detecting and determining the presence of the complex as a measurement for the presence or the amount of the biopolymer target contained in the sample.

A further aspect of the present invention provides a device comprising a plurality of unmodified biopolymer and a solid support. The solid support has at least one surface comprising pendant acyl fluoride functionalities, and the biopolymer is attached to the solid support by reaction with the pendant acyl fluoride functionalities.

The present invention is well-suited for use in creating polynucleotide arrays, such as genosensor arrays and other array-based systems such as differential gene expression micro-arrays. The polynucleotide arrays may be used for the evaluation or identification of biological activity. The present invention may also be used in creating polynucleotide arrays for the purpose of polynucleotide sequencing. Further, the present invention may be used in hybridization assays and immunoassays.

The present invention provides many advantages. It allows for the attachment of unmodified biopolymers directly to a solid support. It thus simplifies and further increases the versatility of a process for creating biopolymer arrays.

There is both an economic advantage as well as a technical advantage to this invention.

5 First, costly production of modified biopolymers, such as amino-modified DNA, may be avoided. Post-modification processing of oligonucleotides is time-consuming and can substantially increase costs by as much as two-fold. Second, the task of making arrays is greatly simplified, since post-modification processes are no longer required.

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Description of the Figures

10 The above-mentioned and other features of this invention and the manner of obtaining them will become more apparent, and will best be understood, by reference to the following description, taken in conjunction with the accompanying drawings. These drawings depict only a typical embodiment of the invention and do not therefore limit its scope. They serve to add specificity and detail, in which:

15 Fig. 1a shows the hybridization results of labeled target cDNA from actin, G3PDH, p53 and TNF with their corresponding solid-phase primers. Fig. 1B shows the hybridization results of labeled cDNA targets with their corresponding probe cDNA immobilized by the same process.

Fig. 2 shows an array pattern created by manual pipettor spotting.

20 Fig. 3 shows the hybridization results of cDNA targets with manually spotted arrays of unmodified oligonucleotide primer pairs.

Fig. 4 shows the hybridization results of the biotinylated Fib target oligonucleotide with both amino-modified and unmodified complementary probes of the array.

25 Fig. 5 is an image analysis of the mean signal intensities for each probe spot within the array.

Fig. 6 shows Biomek HDRT printing of cDNA and BAPA markers onto acyl fluoride activated polypropylene films.

Fig. 7 shows the stability of polypropylene-based cDNA arrays.

Fig. 8 shows the immobilization of Protein A.

30 Fig. 9 shows the immobilization of streptavidin.

Fig. 10 shows the immobilization of Gt anti-biotin IgG antibody.

Detailed Description of the Invention

One aspect of the present invention provides a method of attaching unmodified biopolymers to a solid support. The method includes the steps of:

- (a) providing unmodified biopolymers;
- (b) providing a solid support having at least one surface comprising pendant acyl fluoride functionalities; and
- (c) contacting the unmodified biopolymers with the solid support under a condition sufficient for allowing the attachment of the biopolymers to the solid support.

The term "biopolymer" as used herein refers to nucleic acids, polynucleotides, polypeptides, proteins, carbohydrates, lipids and analogues thereof. As used herein, "polynucleotide" refers to a polymer of deoxyribonucleotides or ribonucleotides, in the form of a separate fragment or as a component of a larger construction. "Polynucleotide" as used herein may be DNA, RNA, a DNA analog such as PNA (peptide nucleic acid), or a synthesized oligonucleotide. The DNA may be a single or double-strand DNA, or a DNA amplified by PCR technique. The RNA may be a mRNA. The length of the polynucleotides may be 3 bp to 10 kb. In accordance with one embodiment of the present invention, the length of a polynucleotide is in the range of about 50 bp to 10 Kb, preferably, 100 bp to 1.5 kb. In accordance with another embodiment of the present invention, the length of a synthesized oligonucleotide is in the range of about 3 to 100 nucleotides. In accordance with a further embodiment of the present invention, the length of the oligonucleotide is in the range of about 15 to 20 nucleotides.

As used herein, "polypeptide" refers to a polymer of amino acids, wherein the α -carboxyl group of one amino acid is joined to the α -amino group of another amino acid by a peptide bond. A protein may comprise one or multiple polypeptides linked together by disulfid bonds. Examples of the protein include, but are not limited to, antibodies, antigens, ligands, receptors, etc.

It is a discovery of the present invention that biopolymers of the present invention may be attached to a solid support without any modification to the biopolymers. For example, in accordance with one embodiment of the present invention, biopolymers such as, but not limited to, polynucleotides, protein A, antibodies or streptavidin may be attached to a solid support without any modifications to the biopolymers.

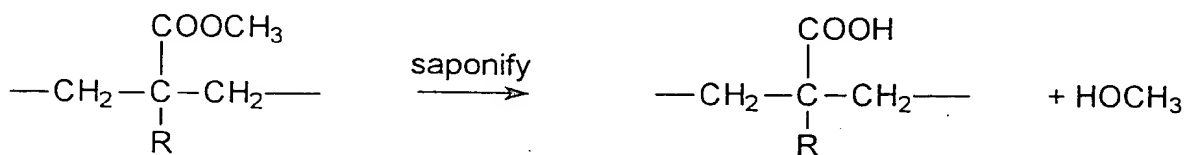
For the purpose of the present invention, the solid support of the present invention may be any material capable of being derivatized to form acyl fluoride functionalities on at least one surface of the solid support. Examples of a solid support include, but are not limited to, polymeric materials, glasses, ceramics, natural fibers, silicones, metals and composites thereof.

In accordance with one embodiment of the present invention, a solid support of the present invention may be fabricated of a polymeric material having at least one surface with pendant acyl fluoride functionalities. The detailed description of the solid support and the methods of making the solid support are described in the pending U.S. application serial number 08/797,222, entitled "Polymeric Reagents for Immobilizing Biopolymers," the relevant content of which is incorporated herein in its entirety by reference.

Briefly, polymeric materials suitable for fabricating solid supports can be any material capable of being derivatized to form acyl fluoride functionalities on at least one surface of the solid support. For example, polymeric materials with pendant carboxyl functionalities or polymeric materials capable of being modified to support carboxyl groups can be reacted with suitable reagents to form acyl fluoride functionalities. In one embodiment of the present invention, solid supports are fabricated of ethylene acrylic acid copolymers, ethylene methacrylic acid copolymers, or derivatized polypropylene. Those skilled in the art will recognize that polymeric materials capable of being derivatized to support carboxyl groups which in turn can be modified to provide surface acid fluoride functionalities include a wide range of materials. For example, aminated polypropylene reacted with a cyclic anhydride, e.g., succinic anhydride, to provide carboxyl groups suitable for converting to acyl fluoride is particularly useful.

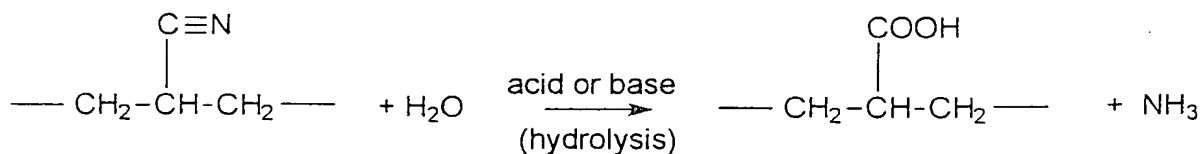
Additional suitable polymeric materials include methylmethacrylate or methylacrylate saponified to expose a pendant carboxyl group:

Methyl(meth)acrylate



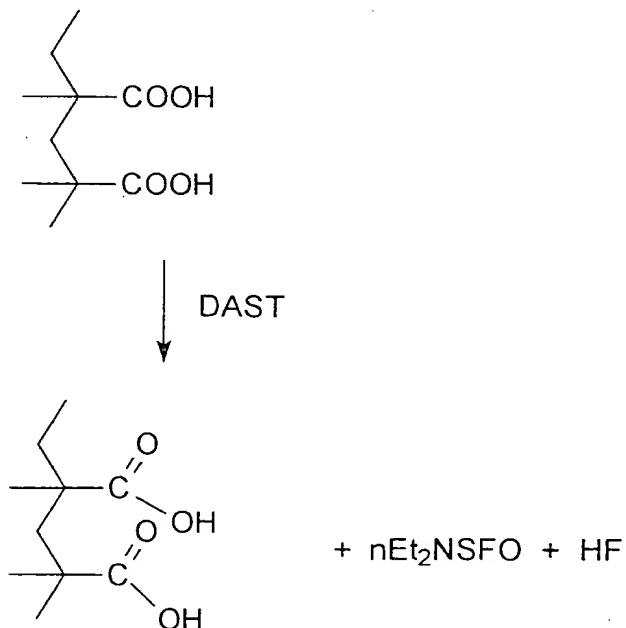
Still other polymeric materials easily derivatized to provide a reactive carboxyl group are hydrolyzed polyacrylonitrile or hydrolyzed polymethacrylonitrile:

(Meth)acrylonitrile

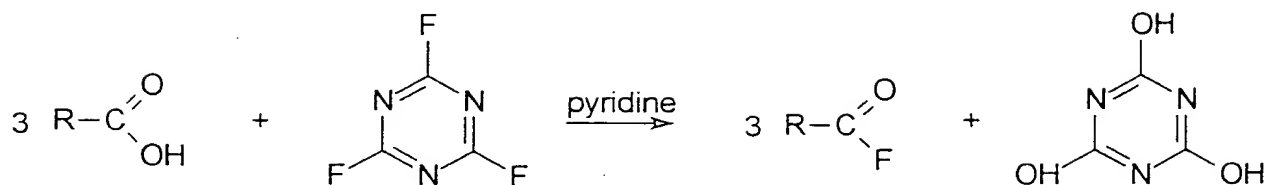


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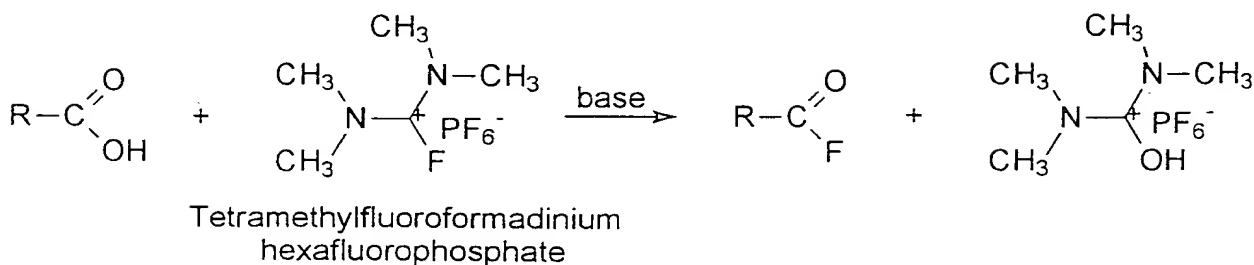
In accordance with the present invention, suitable reagents for forming acyl fluoride functionalities on at least one surface of the solid support broadly include carboxyl reactive fluorinating reagents. A most preferred reagent is (diethylaminosulphur) trifluoride (DAST) which reacts with pendant carboxyl groups in the following reaction:



Other reagents include cyanuric fluoride:



and tetramethylfluoroformadinium hexafluorophosphate:



In a most preferred embodiment of the present invention, ethylene acrylic acid copolymers or ethylene methacrylic acid copolymers formed into sheets having a thickness of about 0.5 mm are modified on at least one surface by exposing the surface to about a 5% solution of DAST for a period of several hours. After stopping the reaction, using dichloromethane and acetonitrile washes, the film is ready for use in immobilizing or synthesizing biopolymers as described below. Advantageously, it has been discovered that acyl fluoride activated polymeric materials are surprisingly stable at ambient conditions and, when stored in a cool dry environment, have unlimited shelf lives.

The solid supports of the present invention may also be prepared by methods comprising the steps of providing a solid support fabricated of ethylene acrylic acid copolymer or ethylene methacrylic acid copolymer and derivatizing at least one surface of the solid support by reacting the surface with an activating agent. Suitable activating agents are reagents capable of reacting with the acrylic or methacrylic carboxyl group to form reactive pendant functional groups, e.g., active acyl functionalities.

In accordance with another embodiment of the present invention, a solid support of the present invention may also be prepared by coating of an inert solid support with a polymeric

material containing pendant acyl fluoride functionalities. For example, poly-glutamic acid carboxyl groups may be converted to acyl fluoride by a suitable fluorinating agent, such as DAST. The converted polymeric material may be used to coat a microtiter plate or other solid supports.

5 Because the solid support of the present invention is particularly useful in the preparation of biopolymer arrays for the evaluation or identification of biological activity, the solid support is preferably in the form of a device having at least one flat planar surface. The size of the solid support can vary and depends upon the final use of the immobilized biopolymers. Those skilled in the art will appreciate that arrays of biopolymers immobilized on miniaturized solid supports have been under development for many years. These solid supports can be measured in terms of mm^2 and can have numerous different immobilized biopolymers, each with different biopolymers attached to a different site specific location on the miniaturized solid support. Solid supports in the form of dip sticks are also within the scope of the present invention. As known in the art, dip sticks typically are rectangular in shape with each side measuring a few centimeters. On the other hand, large biopolymers such as polynucleotide arrays, utilized for sequencing whole genomes, may have dimensions measuring a meter or more.

10 In order to accommodate a number of different testing techniques including specialized testing equipment, suitable solid supports can also be molded into any of a variety of shapes.

15 For example, it may be advantageous to mold a biopolymer array holder of the same polymeric material utilized to fabricate the solid support. In such a system the holder is the solid support and may be any shape, including one which is easily handled by an automated diagnostic system in which robotic arms move biopolymer arrays between reaction stations and detection stations. Preferably, when such a holder is the solid support, it incorporates a planar or continuous surface suitable for attaching biopolymers.

20 In accordance with one embodiment of the present invention, a solid support of the present invention may be made of a porous or non-porous material. In accordance with another embodiment of the present invention, a solid support of the present invention may be in a form of threads, sheets, films, gels, membranes, beads, plates and like structures. In accordance with a further embodiment of the present invention, a solid support may be fabricated from plastic in the form of a planar device having discrete isolated areas in the form of wells, troughs, pedestals, hydrophobic or hydrophilic patches, diecut adhesive reservoirs or

laminated gaskets diecuts that form wells or other physical barriers to fluid flow. Examples of such a solid support include, but are not limited to, a microplate or the like.

The biopolymers of the present invention are attached to a solid support of the present invention by contacting the unmodified biopolymers with the solid support under a condition
5 sufficient for allowing the attachment of the biopolymers to the solid support. A condition is sufficient if it allows the unmodified biopolymers to react with acyl fluoride of a solid support for covalently attaching the biopolymers to the solid support. While not wanting to be bound by the theory, it is believed that, under the conditions of the present invention, unmodified biopolymers may be attached to a solid support by displacement of the fluoride group
10 contained in the solid support with a nucleophile of a biopolymer.

In accordance with one embodiment of the present invention, the step of contacting the acyl fluoride activated surface of the solid support under conditions which cause unmodified polynucleotides to react with acyl fluoride is accomplished by exposing the solid support
15 surface to the unmodified polynucleotides in the presence of an aqueous buffer, preferably with a neutral or basic pH. Bringing the acyl fluoride functionalities into contact with the unmodified polynucleotides under neutral or basic pH conditions results in the attachment of the polynucleotides directly to the surface of the solid support. For the purpose of the present invention, a basic pH condition is a condition that has a pH greater than 8. A basic pH condition is sufficient if it allows the attachment of the polynucleotides to the solid support. In
20 accordance with one embodiment of the present invention, the basic pH condition of the present invention has a pH of about 9 to 12. It should be understood that the basic pH condition may vary, depending on the method used. One skilled in the art can readily ascertain the basic pH condition of a particular attachment reaction in view of the disclosure of the present invention.

A solid support of the present invention may be contacted by unmodified biopolymers
25 by methods that are known in the art. For examples, the contacting step may be carried out by jet printing, solid or open capillary device contact printing, microfluidic channel printing, silk screening, and a technique using printing devices based upon electrochemical or electromagnetic forces. Alternatively, the contacting step may be carried out by spotting the
30 unmodified biopolymers to the solid support.

For example, in one embodiment the solid support is exposed to unmodified polynucleotides by manual spotting. Examples of manual spotting include, but are not limited

to, manual spotting with a pipettor or with a Biomek pin tool. In this case, the preferred aqueous base may be sodium bicarbonate-carbonate with a pH in the range of 9 to 10. In another embodiment, the solid support is exposed to unmodified polynucleotides by jet printing techniques. Thermal inkjet printing techniques utilizing commercially available jet printers and piezoelectric microjet printing techniques as described in U.S. Patent No. 4,877,745 can be utilized to spot unmodified polynucleotides to solid supports. In this case, the aqueous base may be a LiCl salt solution with a pH of about 10 to 12.

It should be understood that the solid support of the present invention may be exposed to biopolymers by any methods as long as the biopolymers are put in contact with the solid support. It should also be understood that other aqueous buffer systems, which are not explicitly described here, may also be used in the present invention as long as the buffer system provides a sufficient condition that allows the attachment of biopolymers to the solid support once they are in contact with each other.

In accordance with embodiments of the present invention, the concentration of unmodified biopolymers contained in aqueous solutions may vary, depending on the type of molecule, the molecule size, the molecule structure, and other factors that may influence solubility of the molecules. For example, when the attached polymers are polynucleotides, preferably they are in the range of 5 nM to 40 :M. More preferably, they are in the range of 5 nM to 5 :M.

In accordance with the present invention, it is preferable, subsequent to the attachment of biopolymers to solid supports, to "block" the nonreacted acyl fluoride functionalities from further unwanted reactions. For the purpose of the present invention, the residual acyl fluoride groups contained in the solid support may be blocked by any chemicals that can inactivate the remaining surface reactive groups. For example, unreacted acyl fluoride functionalities may be reacted with ammonium hydroxide to form carboxamide or with ethanol to form esters. Those skilled in the art will recognize, however, that a host of blocking reactions are possible.

As mentioned above, many applications for utilizing immobilized biopolymers require that biopolymers be immobilized at site specific locations on a solid support surface. In order to prepare ordered arrays of biopolymers, including grids and 1 x n arrays of immobilized biopolymers with each biopolymer located at site specific locations, a preselected site on the surface of the activated polymeric material is exposed to a solution of the desired unmodified

biopolymers. In accordance with the present invention, this can be accomplished manually by applying an amount of unmodified biopolymer solution to a preselected location on the solid support. Alternatively, thermal inkjet printing techniques utilizing commercially available jet printers and piezoelectric microjet printing techniques as described in U.S. Patent No.

4,877,745 can be utilized to spot selected solid support surface sites with selected unmodified biopolymers.

In accordance with one embodiment of the present invention, at least 1 to about 1536 different unmodified biopolymers may be attached to at least 1 to about 1536 discrete isolated areas in a solid support of the present invention

Attachment of unmodified biopolymers to acyl fluoride activated solid supports is well-suited for use in the construction of genosensors and other array-based systems such as differential gene expression micro-arrays. A solid support with attached unmodified biopolymers of the present invention may also be used as a device for performing a ligand binding assay or for performing a hybridization assay by either reverse hybridization (probes attached) or southern blot (target attached). Such a device may also be used in an immunoassay.

Accordingly, one aspect of the present invention provides a method for analyzing a biopolymer target in a sample. The method comprises the steps of:

- (a) providing a solid support fabricated of a material having pendant acyl fluoride groups on at least one surface;
- (b) providing an agent that can form a complex with the biopolymer target, wherein the agent comprises a second biopolymer;
- (c) contacting the solid support with either the agent or the biopolymer target under a condition that allows the attachment of either the unmodified agent or the unmodified biopolymer to the solid support, wherein the agent and the biopolymer target are unmodified;
- (d) contacting the solid support attached with the unmodified agent with the biopolymer target, or contacting the solid support with the attached, unmodified biopolymer target with the agent under a condition that allows the formation of a complex comprising the agent and the biopolymer target;
- (e) detecting and determining the presence of the complex as a measurement for the presence or the amount of the biopolymer target contained in the sample.

For the purpose of the present invention, either the target biopolymer or the agent for the target biopolymer may be bound to a solid support of the present invention. For examples, in southern blot or northern blot, target biopolymers are first bound to a solid support.

Probes, preferably labeled, are used to contact the solid support to detect the existence of the target biopolymers. To the contrary, in ligand binding assays or affinity purification assays, probes are preferably bound to a solid support first.

For the purpose of the present invention, an agent of the present invention comprises a biopolymer that can recognize the target biopolymer and bind to the target biopolymer. An agent may comprise a biopolymer selected from a group consisting of nucleic acids, polypeptides, proteins, carbohydrates, lipids and analogues thereof. For example, when the target biopolymer is a polynucleotide, the agent may comprise a polynucleotide that is complimentary to the target polynucleotide. When the target biopolymer is a receptor or a ligand, the agent may comprise a ligand or a receptor that respectively recognizes the biopolymer and binds to the biopolymer. When the target biopolymer is an antigen, the agent may comprise an antibody that recognizes the antigen, or vice versa. In addition, the target biopolymers may be labeled with a binding agent. In this case, the agent may comprise a binding agent that recognizes the labeled biopolymers. For example, a biopolymer of the present invention may be labeled with biotin. In this case, the agent may comprise streptavidin which can bind to biotin of the target biopolymer.

For the purpose of the present invention, a condition is sufficient if the agent can bind to the target biopolymer to form a complex. Such a condition may vary, depending on the type of molecules and the type of bindings. One skilled in the art can readily determine the suitable conditions for binding in view of the teaching of the present invention.

In accordance with embodiments of the present invention, both the target biopolymers or agents of the present invention may be labeled with a reporter molecule. Examples of reporter molecules include, but are not limited to, dyes, chemiluminescent compounds, enzymes, fluorescent compounds, metal complexes, magnetic particles, biotin, haptens, radio frequency transmitters, and radioluminescent compounds. One skilled in the art can readily determine the type of reporter molecule to be used once the type of target biopolymers is determined.

Another aspect of the present invention also provides a device for performing hybridization assays, immunoassay or other assays. A device of the present invention

comprises a plurality of unmodified biopolymers and a solid support. The solid support has at least one surface comprising pendant acyl fluoride functionalities, and the biopolymer is attached to the solid support by reaction with the pendant acyl fluoride functionalities.

For the purpose of the present invention, the attached biopolymers may be the same or different. If the biopolymers are different, preferably they are located in discrete, isolated areas of the solid support to form arrays. For example, a solid support may be a microplate. Different biopolymers may be attached to different wells of the microplate for forming arrays. In accordance with one embodiment of the present invention, at least 1 to 1536 unmodified biopolymers, such as probes, may be attached to at least 1 to 1536 wells of a microplate.

The solid support of the device, such as a microplate, may be surface treated with acyl fluoride functionalities, and then biopolymers may be attached to the solid support by reaction with the pendant acyl fluoride functionalities. Alternatively, biopolymers of the present invention may be printed onto the surface of a plastic disk containing pendant acyl fluoride functionalities, and the disk is then inserted into the bottom of the microplate well.

The following examples are intended to illustrate, but not to limit, the scope of the invention. While such examples are typical of those that might be used, other procedures known to those skilled in the art may alternatively be utilized. Indeed, those of ordinary skill in the art can readily envision and produce further embodiments, based on the teachings herein, without undue experimentation.

Example 1

Hybridization of cDNA to Printed Arrays of Unmodified Polynucleotides

Printing of Primer Arrays

Unmodified oligonucleotide primers used for amplification of actin, G3PDH, p53 and TNF alpha genes were prepared by dilution into saturated LiCl, pH 12 stock solution at approximately saturated LiCl: water (60:25 v/v). The resulting printing ink was at a final concentration of 10 μ M oligonucleotide. Using a BioDot Dispenser System, primers were deposited by jetting 12.5 nL of each ink onto a molded piece (Biotip) of acyl fluoride activated ethylene methacrylic acid copolymer (EMA) to form an array. The 9 X 8 array consisted of 9 replicate spots for each 3' & 5' primer pair for the 4 genes ($9 \times 2 \times 4 = 72$ spots). Following printing, the arrays were dried overnight under vacuum and remaining surface reactive groups inactivated (blocked) for two hours in ethanol. The arrays were rinsed briefly with water in preparation for use in hybridization.

Printing of cDNA Probe Arrays

In a similar manner, double-stranded DNA probes were printed onto acyl fluoride activated substrates. The cDNA was amplified by PCR of 1st strand DNA without incorporation of biotin label. The cDNA PCR products (cDNA amplicons) were purified using gel filtration spin-columns (Xtreme, Pierce Chemical) to remove primers, dNTPs, along with cofactors and Taq enzyme. The amplicons were eluted from the columns with deionized water and prepared for printing by dilution into 1M LiCl, pH 12 to yield an ink of LiCl: water (1:1 v/v) at a final cDNA concentration of 5 nM. The same spotting procedure was used as described for primer printing, except in this case the array consisted of nine replicates of six genes (9 X 6 = 54 spots): actin cDNA, beta microglobulin cDNA, G3PDH cDNA, p53 cDNA, transferrin cDNA and TNF alpha cDNA. The droplet size dispensed was approximately 16 nL.

Hybridization

Biotips with cDNA arrays attached were denatured for 15 minutes in 200 µL denaturant (0.15 M NaCl, 0.5M NaOH), then rinsed in stringency buffer (2X SSC, 0.01% SDS, pH 7.0) just prior to hybridization. In the case of Primer Array Biotips, no denaturation step was used. Biotin-labeled PCR products of actin, G3PDH, p53 and TNF alpha derived from a 1st strand cDNA pool (liver) were prepared for hybridization to the Primer Arrays or cDNA Arrays as follows: 10 µL PCR solution was diluted with 10 µL water and 50 µL denaturant added. The solution was incubated for 10 minutes at ambient temperature followed by the addition of 150 µL of neutralization buffer (0.3M Tris, pH 7.5, 2.4xSSC, 0.02% SDS). After mixing, the solution was placed in well of a 24-well polypropylene cell culture plate and the Biotip immersed. Hybridization was allowed to proceed for 60 minutes at 25°C for cDNA Arrays of 60°C, 60 minutes for Primer Arrays with shaking in a humidified chamber. The Biotips were then removed from the hybridization solution and placed in another well containing 2 X SSC, 0.01% SDS for a stringency rinse (at the same temperature used for hybridization) for 10 minutes. Upon a final rinse in the above stringency buffer, the Biotip was blotted to remove excess solution and placed in streptavidin-alkaline phosphatase conjugate for 30 minutes at ambient temperature. Following extensive rinsing in stringency buffer, the Biotip was again blotted, then placed in ELF reagent (fluorescent substrate for alkaline phosphatase, Molecular Probes, Inc.) for signal development. The signal was allowed

to develop for 30 minutes. Following a brief water rinse, the array signal was read using a CCD camera system.

Results

As shown in Fig. 1a, labeled target cDNA from actin and G3PDH hybridized specifically to their corresponding solid-phase primers, while cDNA from p53 showed cross-hybridization to G3PDH primers. TNF alpha also cross-hybridized to 3' primers of actin and p53. These results are similar to those using labeled cDNA targets hybridized to probe cDNA immobilized by the same process (Fig. 1b)

Example 2

Hybridization of cDNA Targets to Manually Spotted Arrays of Unmodified

Oligonucleotide Primer Pairs

Manual Spotting of Primer Arrays

Oligonucleotide primers for the amplification of actin, p53 and TNF alpha genes were prepared at the following concentrations for spotting in 0.5 M sodium bicarbonate buffer, pH 9:

Primer	μM (final conc.)
Actin 3'	5.01
Actin 5'	5.00
P53 3'	3.73
P53 5'	3.83
TNF 3'	3.80
TNF 5'	3.70

Approximately 1 μL drop of each of these were spotted onto an acyl fluoride activated plastic support (EMA) using a manual pipettor creating an array similar to the pattern shown below in Fig. 2.

After spotting, the arrays were incubated at 25°C, 1 hour in a humidified chamber. They were then removed from the chamber and allowed to air-dry 25 minutes at ambient temperature, followed by an additional drying at 30°C for 10 minutes. The plastic supports were then immersed in ethanol for 60 minutes to block residual reactive surface groups, followed by a final 10- minute soak in deionized water.

Hybridization

Biotin-labeled PCR products of actin, p53 and TNF alpha derived from a 1st strand cDNA pool (liver) were prepared for hybridization to the Primer Arrays as follows: 10 µL PCR solution was diluted with 10 µL water and 30 µL denaturant added. The solution was incubated for 10 minutes at ambient temperature followed by the addition of 150 µL of neutralization buffer. After mixing, the solution was placed in a well of a 24-well polypropylene cell culture plate and the Biotip immersed. Hybridization was allowed to proceed for 60 minutes at 60°C with shaking in a humidified chamber. The Biotip was then removed from the hybridization solution and placed in another well containing 2 X SSC, 0.01% SDS for a stringency rinse at 60°C for 10 minutes. Upon a final rinse in the above stringency buffer, the Biotip was blotted to remove excess solution and placed in streptavidin-alkaline phosphatase conjugate for 30 minutes at ambient temperature. Following extensive rinsing in stringency buffer, the Biotip was again blotted, then placed in ELF reagent for signal development. The signal was allowed to develop for 30 minutes. Following a brief water rinse, the array signal was read using a CCD camera system.

Results

Fig. 3 shows the results of the hybridization. The biotin-labeled cDNA targets hybridized specifically to their complementary immobilized primer probes. Actin and p53 targets showed little cross-hybridization to non-complementary primer probes tethered to the surface. However, TNF target exhibited significant cross-hybridization, especially to the 3' primers.

Example 3

Hybridization of Biotin-labeled Oligonucleotide Targets to Printed Arrays of Amino Modified and Unmodified Oligonucleotide Probes

Printing of Fib Probe Arrays

5'-amino modified (Fib-a) and unmodified oligonucleotide (Fib) probes were prepared by dilution into saturated LiCl, pH 12 to yield saturated LiCl: water (60:25 v/v) printing ink at final concentrations from 10 µM to 40 µM for Fib; and 10 µM for Fib-a. Using a BioDot Dispenser System, nine replicate spots of each probe solution were deposited by jetting 16 nL of each ink onto a molded piece (Biotip) of acyl fluoride activated EMA to form an array.

Following printing, the arrays were dried overnight under vacuum and remaining surface

Sub B1
 reactive groups inactivated (quenched) for two hours in ethanol. The arrays were briefly rinsed with water in preparation for use in hybridization.

Fib Probe: 5'-CGGCTGGACACGCTTCTGTAG-3'

Fib-a Probe: 5'-NH₂-CGGCTGGACACGCTTCTGTAG-3'

Fib Target: 5'-Biotin-CTACAGAAGCGTGTCCAGCCG-3'

Hybridization

Fib target (100 nM, final conc.) was prepared by mixing 20 μ L of a 1 μ M stock solution with 30 μ L of denaturant. The target was held under denaturing conditions for 10 minutes at ambient temperature followed by the addition of 150 μ L of neutralization buffer. The Biotip was immersed in the solution held in a well of a 24-well polypropylene microtiter plate which was in turn placed in a humidified chamber for 60 minutes at 25 C. Following a 10-minute rinse in 2X SSC, 0.01% SDS, pH 7.0 stringency buffer, the Biotip was incubated for 30 minutes at ambient temperature in a streptavidin-alkaline phosphatase conjugate solution prepared in stringency buffer. The Biotip was rinsed 2 X with stringency buffer to remove unbound conjugate; and then immersed in ELF reagent for signal development for 30 minutes. Following a brief water rinse, the Biotip array signal was read using a CCD camera system.

Results

Fig. 4 shows that the biotinylated Fib target oligonucleotide hybridized to both amino-modified and unmodified complementary probes of the array.

Image analysis of the mean signal intensities for each probe spot within the array was conducted for three separate Biotip arrays. Fig. 5 shows that amino-modified probe and unmodified probe hybridizations were of approximately the same level when considering the range of values estimated from the standard deviation.

Example 4

Preparation and Characteristics of cDNA Arrayed onto Acyl Fluoride Activated

Polypropylene Films

Activation of Polypropylene Films

Polypropylene film was surface aminated by radio frequency plasma (U.S. Patent No. 5,554,501) and amine functionalities converted to carboxyl groups by reaction with succinic anhydride. The acyl fluoride activation was accomplished using the DAST reagent as

described previously in the copending U.S Patent application serial number 08/797,222. Acyl fluoride activated polypropylene film was stored dry under argon at -20 C until needed.

Printing of cDNA Arrays

Unmodified cDNA from pcr amplification of 1st strand cDNA (liver tumor) was purified by gel filtration spin column (XTreme, Pierce Chemical) to remove primers, dNTPs, cofactors and Taq enzyme. The purified cDNA was eluted from the columns with water, then diluted into 0.5 M sodium bicarbonate buffer, pH 9 for coupling. A Biomek 2000™ robotic system equipped with a 384-pin HDRT system was used to print cDNA solutions onto the substrate. A set of BAPA markers, 5-(Biotinamido)pentylamine (Pierce Chemical), were also printed at both ends of the film, thereby flanking the tethered cDNAs. BAPA, which binds streptavidin-enzyme conjugate independently of hybridization, serves as an internal control for assay robustness. Following printing, the films were dried at 35 C for 15 minutes and then immersed in ethanol for two hours to block residual surface reactive groups. The films were briefly rinsed in deionized water and allowed to air dry. The 8 cm X 12 cm film was sectioned into 12 pieces with each strip containing two copies of a 3 X 3 replicate of each cDNA along with six copies of 3 X 3 replicates of the BAPA marker. The resulting cDNA Array film strips were stored dry at -20°C or room temperature prior to hybridization.

Hybridization

For hybridization, each film strip was denatured for 15 minutes in 150 µL of denaturant dispensed onto a glass microscope slide. Following the denaturation step, the film strips were removed from the slide and briefly rinsed in stringency buffer to remove residual denaturant. A mixture of biotin-labeled pcr products of actin, G3PDH and TNF alpha derived from a 1st strand cDNA pool (liver tumor) were prepared for hybridization in a 15 mL polypropylene tube as follows: 15 µL of each cDNA pcr solution was mixed and then diluted with 180 µL water. Then 337.5 µL of denaturant was added. Denaturation was allowed to proceed for 10 minutes at ambient temperature followed by the addition of 1687.5 µL of neutralization buffer to the tube. A 150 µL aliquot was transferred onto glass microscope slides and the film arrays placed over the solution with the DNA side down. Hybridization was allowed to proceed for 60 minutes at 60 C with shaking in a humidified chamber. The film strips were then removed from the hybridization solution, transferred to 50 mL polypropylene screw cap culture tubes and the films first rinsed briefly, then immersed in 40 mL of 2 X SSC, 0.01% SDS buffer preheated to 60 C for a stringency rinse at 60 C for 10

minutes. Upon a final rinse in the above stringency buffer, the films were blotted to remove excess solution and placed on glass microscope slides containing a solution of streptavidin-alkaline phosphatase conjugate for 30 minutes at ambient temperature. Following extensive rinsing in stringency buffer, the films were again blotted, then placed in ELF reagent for signal development. The signal was allowed to develop for 30 minutes. Following a brief water rinse, the array signal was read using a CCD camera system.

Results

An array of cDNA and BAPA markers was successfully printed onto acyl activated polypropylene films as determined by hybridization of a mixture of the corresponding cDNAs (Fig. 6). Fig. 6 shows Biomek HDRT printing of cDNA and BAPA markers onto acyl fluoride activated polypropylene films (day 7 hybridization to 3 separate film strips). Storage of film arrays, either at ambient temperature or at -20°C , did not adversely affect hybridization over an approximate three-month period (Fig. 7). Fig. 7 shows the stability of polypropylene-based cDNA arrays. Signal intensities were normalized to the BAPA marker intensities to account for differences in assay conditions over the extended period. All cDNA probes maintained hybridization efficiency under either temperature storage condition. The TNF signal was consistently lower than the BAPA marker signals, while the remaining cDNA hybridization signals remained approximately at the same intensity as the BAPA markers.

Example 5

Protein Immobilizations

Proteins, Buffers and other reagents

1. Biotinylated Alkaline Phosphatase: Pierce Immunopure® (lot # 95120774)
2. Rabbit anti-goat IgG (H+L): Zymed Laboratories, South San Francisco, CA (lot # 2112098)
3. Goat anti-biotin IgG: Sigma Chemical Co. (lot # 116H8842), affinity isolated antibody; reconstituted to 1 mg/mL in PBS.
4. Protein A: Zymed Laboratories, South San Francisco, CA (lot # 21212416); reconstituted 5 mg in 2 mL deionized water (2.5 mg/mL).
5. Streptavidin (from *S. avidinii*): Zymed Laboratories, South San Francisco, CA (lot # 70536128); reconstituted 5 mg in 1 mL deionized water (5 mg/mL).

6. Casein: Hammersten (lot # BDH44020)
7. Coupling buffer: 1 M sodium carbonate, pH 9 or 10 diluted to 0.8 M with protein solution
8. ELF[®] Reagent (ELF[®]-97 Endogenous Phosphatase Detection Kit): Molecular Probes, Eugene, OR (lot # 6771)
- 5 9. High salt buffer: 1.0 M Tris, 1.5 M NaCl, pH 7.5 (Digene Neutralizing Buffer); Digene Diagnostics, Beltsville, MD (lot # 0094MX95)
10. Quenching/Blocking buffer: 1 mg/mL casein, 50 mM Carbonate, 0.15 M NaCl, pH 8.5
11. TBS: 50 mM Tris, 0.15 M NaCl, pH 7.5

10 **Methods**

Immobilization of Proteins on Biotips

Proteins (Protein A, streptavidin, rabbit anti-goat IgG) were diluted into 1 M carbonate buffer, pH 9 or 10, to a concentration of 0.5-1.0 mg/mL. The protein solution was applied to the surface of acyl fluoride activated plastic Biotips in 0.5 μ L drops. The tips were incubated in a high humidity chamber at 25°C for 1 hr. They were then removed from the chamber and allowed to air dry completely (~15 min) before being placed in the quenching/blocking solution. The tips were quenched by placing in 1.0 mL solution in a 24-well plate and vortexing vigorously for a minimum of 30 min. After quenching, the tips were rinsed with deionized water for 10 min with shaking and air dried.

Detection of Alkaline Phosphatase with ELF Reagent

For each tip, 50 μ L ELF reagent is prepared according to the Molecular Probes procedure (i.e. 19 parts Reagent B plus 1 part Reagent A) and applied to the surface of the tips. The tips with ELF reagent were incubated for 30 min in a box partially filled with water to maintain humidity. Afterwards, the tips are rinsed three times with deionized water. The tips are photographed under UV light at 365 nm excitation with a 520 nm lens filter. Exposure time for the CCD camera was typically 10 sec.

Experimental Results

Immobilization of Protein A on Biotips

5 Signal Development & Detection:

Incubated Rb anti-Gt IgG, 1:100 in TBS for 1 hr, followed by Gt anti-biotin IgG in TBS for 1 hr, and then biotin-AP conjugate in 2X SSC, 0.01% SDS for 1 hr. Image developed with ELF. The immobilized Protein A was demonstrated to be functional in it's capture of the rabbit (Rb) antibody in the above sandwich-based immunoassay.

Immobilization of streptavidin on Biotips

Prepared at 1 mg/mL in 0.8 M carbonate, pH 9. Incubated for 1 hr at room temperature. Quenched for 45 min on Vortexer with different blocking agents (at 1 mg/mL).

15 Signal Development & Detection:

Incubated biotinylated alkaline phosphatase diluted 1:100 in 2X SSC, 0.01% SDS buffer for 1 hr. ELF reagent incubation for 30 min. at room temperature. Immobilized streptavidin was demonstrated to be functional in the specific capture of a biotinylated enzyme under a variety of blocking conditions aimed at reducing non-specific binding. (Fig. 9)

Immobilization of Antibody on Biotips

Gt anti-biotin IgG immobilized on Biotips in the following:

- 25 1. 1.0 mg/mL in PBS, pH 7.2 incubated for 30 min
2. 1.0 mg/mL in PBS, pH 7.2 incubated for 60 min
3. 0.5 mg/mL in 0.8 M carbonate, pH 10 incubated for 30 min
4. 0.5 mg/mL in 0.8 M carbonate, pH 10 incubated for 60 min

30 Signal Development & Detection:

Incubation with biotin-alkaline phosphatase conjugated diluted 1:100 in high salt buffer for same time as antibody incubation. Development with ELF reagent for 30 min. at room

temperature. The antibody, goat anti-biotin IgG, was successfully immobilized at both pH 7,2 and pH 10, as demonstrated by specific capture of biotin labeled reporting enzyme (alkaline phosphatase) (Fig. 10).

The foregoing is meant to illustrate, but not to limit, the scope of the invention.

- 5 Indeed, those of ordinary skill in the art can readily envision and produce further embodiments, based on the teachings herein, without undue experimentation.

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